# Effects of storm events on mobilisation and in-stream processing of dissolved organic matter (DOM) in a Welsh peatland catchment

Kari Austnes · Christopher D. Evans · Caroline Eliot-Laize · Pamela S. Naden · Gareth H. Old

Received: 18 November 2008 / Accepted: 2 December 2009 / Published online: 24 December 2009 © Springer Science+Business Media B.V. 2009

**Abstract** Peatlands are important contributors of dissolved organic matter (DOM) to downstream aquatic systems. We investigated the effects of storm events on dissolved organic carbon (DOC) concentrations and DOM quality in a stream draining a Welsh peatland catchment. Intensive stream samples were collected and analysed for pH, DOC, dissolved organic nitrogen (DON), absorbance and fluorescence. Soil water samples and samples of sphagnum pore water

K. Austnes

Department of Plant and Environmental Sciences, Norwegian University of Life Sciences, P.O. Box 5003, 1432 Ås, Norway

K. Austnes (⊠) Norwegian Institute for Water Research, Gaustadalléen 21, 0349 Oslo, Norway e-mail: kari.austnes@niva.no

C. D. Evans Centre for Ecology and Hydrology Bangor, Environment Centre Wales, Deiniol Road, Bangor, Gwynedd LL57 2UW, UK

C. Eliot-Laize · P. S. Naden · G. H. Old Centre for Ecology and Hydrology Wallingford, Maclean Building, Benson Lane, Crowmarsh Gifford, Wallingford, Oxfordshire OX10 8BB, UK

Present Address:
C. Eliot-Laize
Premier Analytical Services, Microscopy,
The Lord Rank Centre, Lincoln Road, High Wycombe,
Buckinghamshire HP12 3QR, UK

were also collected, and a simple end-member mixing model was applied to account for changes occurring during the events. Fluorescence data were interpreted using parallel factor analysis (PARAFAC). DOC concentrations increased and pH decreased during the storm events. The soil water data and the mixing model indicated that this was due to a change of flow paths and draining of the DOC-rich acrotelm. Absorbance indices and the DOC/DON ratio suggested that the DOM released during events was less degraded. There was a striking, inversely related diurnal pattern in absorbance and fluorescence after the discharge peak. The diurnal pattern and a lack of fit with the mixing model suggested that fluorescing DOM was mainly produced in-stream. Fluorescence has been found to peak in the morning and decline during daytime due to photo-bleaching. We hypothesise that the input of additional DOM during events causes a change in the diurnal pattern, giving a peak at mid-day, when the processing of the additional DOM is highest.

**Keywords** DOC · DOM quality · Fluorescence · In-stream processes · Peat stream · Storm events

## Introduction

Peatlands are important contributors of dissolved organic matter (DOM) to downstream aquatic systems. Mean dissolved organic carbon (DOC) fluxes in



rivers draining peat dominated areas are higher than in rivers draining most other landscape types, in the order of 50–100 kg ha<sup>-1</sup> yr<sup>-1</sup> (Aitkenhead and McDowell 2000). Percentage peat cover is found to be a good predictor of DOC concentration across large spatial scales (Aitkenhead et al. 1999).

The concentration of DOC is commonly found to correlate positively with discharge in streams and rivers in temperate and boreal catchments (e.g. Thurman 1985, p. 50; Hope et al. 1994; Soulsby et al. 2003). Likewise, DOC concentrations have frequently been found to increase during storm events (e.g. Hinton et al. 1997; Buffam et al. 2001). However, most studies of discharge-DOC concentration relationships have been conducted in catchments with organo-mineral or mixed soils (Clark et al. 2007). Given the significant contribution of peat systems to DOC concentrations and fluxes, it is important to investigate the relationship between discharge and not only DOC (as a measure of DOM quantity) but also DOM quality in these systems. Studying these relationships is especially important in a climate change perspective, given that annual precipitation, as well as extremes of daily precipitation, are likely to increase in northern Europe (Christensen et al. 2007).

In streams draining organo-mineral soils, the increased DOC concentration at high discharge is commonly explained by changes in flow paths towards increased lateral flow through the upper horizons, where DOC concentrations are higher (e.g. McDowell and Likens 1988; Boyer et al. 1997; Hinton et al. 1998; Inamdar et al. 2006). Change in flow path can be an important control on DOC concentration in peat streams as well, but seems to depend upon the hydrological connectivity between the peat and the underlying bedrock/mineral soil (Clark et al. 2007, 2008). Peat soils consist of an upper horizon (the acrotelm) with roots and decomposing plant material, and a lower horizon (the catotelm) with dense peat (Evans et al. 1999). The stored carbon in the catotelm is hydrologically disconnected from the stream (Billett et al. 2006). Where there is hydrological connectivity between the peat and the mineral soil, base flow is characterised by alkaline DOC poor groundwater (Worrall et al. 2002; Clark et al. 2007). As the water table rises during an event, resulting in subsurface flow in the acrotelm, stream DOC concentration increases due to the input of DOC rich soil water (Evans et al. 1999; Worrall et al. 2002; Soulsby et al. 2003; Clark et al. 2007, 2008). However, further progress of the event may introduce low DOC, rain-like water (Clark et al. 2008), due to exhaustion of the acrotelm or the occurrence of saturated overland flow or macropore flow (Evans et al. 1999; Worrall et al. 2002). Hence, stream concentration can be described by mixing of water from three different source areas, so-called end-members (Worrall et al. 2002). In other peat catchments, base flow is chemically similar to the acidic DOC rich soil water in the acrotelm (Clark et al. 2008). In these systems stream water during events derives from two end-members only: soil water and the rain-like water (Clark et al. 2007). This causes a decrease in stream DOC concentration during events (Clark et al. 2007; Eimers et al. 2008).

Literature on the effects of storm events on DOM quality is sparse. If storm events cause changes of flow paths and not merely dilution, one would assume a change in the quality of the DOM released to the stream (Buffam et al. 2001). DO<sup>14</sup>C data from peat moorland in the upper Conwy catchment showed that base flow releases old, soil-derived DOC, whereas high flow releases younger DOC, probably derived from recent plant material (Evans et al. 2007). Probably, a difference in age also implies different degree of degradation, which in turn affects DOM quality (Qualls and Haines 1992; Kalbitz et al. 2003; Saadi et al. 2006). Spectrophotometric measurement of absorbance and fluorescence can give information on DOM quality in terms of chemical characteristics of the organic material and its bioavailability (e.g. Senesi 1990; Peuravuori and Pihlaja 1997; Croué et al. 1999; McKnight et al. 2001; Kalbitz et al. 2003; Fellman et al. 2008). To our knowledge fluorescence scanning of high frequency event samples has not previously been conducted.

In the present study high frequency measurement of chemical and spectrophotometric characteristics of high frequency stream water samples was conducted in a peatland catchment in Wales. The objectives of the study were: (1) To investigate the effect of storm events on DOC concentration, (2) to investigate the effects of storm events on DOM quality, and (3) to explore the usefulness of fluorescence analyses to assess short term changes in DOM quality.



#### Methods

## Field site

The study site is a small (3.2 km<sup>2</sup>) peatland catchment in north Wales drained by the stream Afon Ddu (Fig. 1). The catchment is a sub-catchment of the upper Conwy catchment, and it is part of the large Migneint blanket peatland area. Catchment altitude ranges from 440 to 540 m.a.s.l., with the main part being below 500 m.a.s.l. Average temperature is 8°C and mean annual rainfall is 2,300 mm. Hydrological response to rainfall events is rapid (Billett et al. 2007). The bedrock is volcanic, mixed rhyolitic and basaltic, of Ordovician origin. Peat soils (histosols) dominate, but at elevated sites podzols occur. The soil depth is usually around 1-2 m, but may be up to 5 m. The vegetation is dominated by heather (Calluna vulgaris L.) and Sphagnum spp. mosses, with scattered grasses and sedges. There is some drainage ditching in the catchment dating from the 1930–1960s.

# Water sampling

Intensive high-flow event sampling in the Afon Ddu was conducted in the autumn of 2007 (sampling point at Ordnance Survey coordinates 277520,344920). Samples were collected during autumn because DOC concentrations were expected to be high (Clark

**Fig. 1** Location of the field site (*left*) and map of the Afon Ddu catchment (*right*)



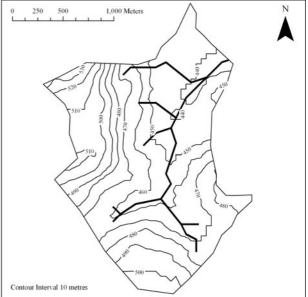
et al. 2008). Samples were collected during three events (September 17–18, October 3–4, October 26–30), hereafter referred to as events 1, 2 and 3. During events 1 and 2, samples were collected for 24 h (event 1 two-hourly, event 2 hourly), while during the larger event three samples were collected two-hourly for 96 h. Samples were collected using a Xian 1000 autosampler (Hach Lange, Germany).

Samples were collected from a number of nearby locations during each event, to provide information on potential end-members. These were sphagnum pore water samples collected by manual squeezing, and three different types of soil water samples collected from blanket peat: bulked soil water collected at 5 cm using micro-rhizon suction samplers and soil water collected at 5 and 10 cm using zero-tension lysimeters.

# Analyses

All samples were split in three for different analyses. The aliquot that was to be analysed for carbon and nitrogen was filtered immediately (within maximum 2 days) through Whatman sterile 0.45  $\mu$ m cellulose nitrate filters. The filters were rinsed with deionised water and an aliquot of sample. All samples were stored at <3°C.

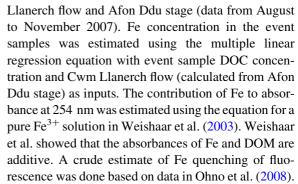
The event sample aliquots that were to be analysed for fluorescence and absorbance were brought to





CEH Wallingford as soon as possible (within 0-5 days). Prior to analysis all samples were filtered through Whatman 1.2 µm glass microfibre GFC filters, after rinsing with water and an aliquot of sample. All equipment used was acid washed. The water used was purified with a NANOpure DIamond Analytical and UV Systems from Barnstead. One cm quartz cells were used. Fluorescence analyses were performed on a Varian Cary Eclipse instrument. Several blanks were run, to ensure that the equipment used had no residual fluorescence. The samples were scanned for emission from 280 to 500 nm at excitation 200 to 400 nm. Absorbance was measured on a Varian Cary 50 instrument, measuring the spectrum from 200 to 800 nm. Whenever the absorbance was above 0.3 cm<sup>-1</sup>, the samples were diluted to below this level, and both analyses repeated, as innerfiltering correction (see below) has been found to be insufficient above this level (Ohno 2002). The sUVa index was calculated as absorbance measured at 254 nm divided by DOC (mg l<sup>-1</sup>) (Vogt et al. 2004) and the E2/E3 index as absorbance measured at 250 nm divided by absorbance measured at 365 nm (Peuravuori and Pihlaja 1997).

Both absorbance and fluorescence may be affected by iron interference. Absorbance may be overestimated, as Fe ions absorb light (Weishaar et al. 2003). Fluorescence may be underestimated, due to quenching of the fluorescence signal caused by Fe-DOM complexation (Zepp et al. 2004; Ohno et al. 2008). To investigate possible iron intereference, data from biweekly sampling (November 2006 to January 2008) at the same spot were used, as Fe concentration was not analysed in the event samples. Fe in the biweekly samples were analysed by ICP-OES. Preliminary analyses showed that the Fe concentration was positively linearly related to DOC concentration  $(R^2 = 0.59)$ . This is in line with Neal et al. (2008), who observed strong correlation between Fe and DOC concentration across a range of UK sites. However, there was also a negative linear relationship with log(flow) (Environment Agency discharge data for the River Conwy at Cwm Llanerch (http:// www.nwl.ac.uk/ih/nrfa/station\_summaries/066/011. html) ( $R^2 = 0.30$ ). Hence, Fe concentration could be reasonably well explained ( $R^2 = 0.78$ ) using multiple linear regression with DOC concentration and log(flow) as explanatory variables. A linear relationship  $(R^2 = 0.82)$  was established between Cwm



The remaining analyses on event samples were conducted at CEH Bangor. Unfiltered samples were analysed for pH using a Metrohm SM 702 Titrino. Filtered samples were analysed for total dissolved nitrogen (TDN) and non purgeable organic carbon (NPOC) by elemental analysis using a Thermalox TOC/TN Analyser. The purging prior to total carbon analysis removes inorganic carbon as CO2 (by addition of 11 µl of 1 M HCl and purging with oxygen for 90 s), so that only dissolved organic carbon (DOC) is determined. Nitrate-N (NO<sub>3</sub>-N) and ammonium-N (NH<sub>4</sub>-N) were analysed by autoanalyser (Skalar SA-40). NO<sub>3</sub>-N was analysed by the sulphanilamide/NEDA/Cd/Cu reduction method, with extinction at 540 nm, and NH<sub>4</sub>-N by the Indolphenol blue method, with extinction at 660 nm. Dissolved organic nitrogen (DON) was calculated by subtracting NO<sub>3</sub>-N and NH<sub>4</sub>-N from TDN.

## Discharge

Discharge at the Afon Ddu sampling site was measured by a Starflow Ultrasonic Doppler instrument with an integrated micrologger (Unidata, Western Australia). Due to instrument failure, the velocity measurements were not correct, so accurate discharges could not be calculated. However stage data were reliable, and provided an effective proxy for the discharge changes associated with each event.

Rain data used were from the Snowdon Environmental Change Network site, about 20 km north-west of the Afon Ddu catchment (Countryside Council for Wales 2008).

# Data analysis

The event data are described quantitatively using different approaches. Base flow composition was



defined as the average stream water composition of samples taken when there was minimal change in stream stage (the first 3, 5 and 11 samples in event 1, 2 and 3, respectively; average for pH calculated via H<sup>+</sup>). In event 1 the stable stage level did not represent a true base flow level, as it closely followed an earlier event. To account for changes in the different parameters during the events, the percentage change from the base flow level to a maximum or minimum level was calculated. Base flow and average soil water (the micro-rhizon and the two zero-tension lysimeter samples) was compared for each event using two sample t-test. Results for sphagnum pore water could not be compared statistically to base flow or soil water, as there was only one sample per event. Correlation of parameters was done using Pearson's r. Results were considered significant for P < 0.05. Minitab Release 14 was used as statistical software. Regression analyses for the Fe-estimation were performed with JMP 7.0.1.

A simple two end-member mixing model was applied to each event and parameter (H<sup>+</sup> and DOC concentration, absorbance at 254 nm and fluorescence intensity), using the average base flow as one end-member and average soil water as the other endmember. This corresponds to the end-member model described by Worrall et al. (2002) but omits the rain end-member (dilution). The model calculates the proportion of each of the two end-members contributing to stream water, starting from the first non-base flow sample. It is a crude model, as there were probably other end-members; base flow is not a true endmember; and none of the parameters are conservative. The intention was simply to assess whether the proposed two end-member system could consistently explain observed variations in different parameters, i.e. that the contribution of the two end-members was consistent for different parameters throughout the event.

Fluorescence data were analysed using parallel factor analysis (PARAFAC) in MATLAB (version R2007b), according to the procedure described by Stedmon and Bro (2008). PARAFAC decomposes the complex set of sample emission-excitation matrices (EEMs) and extracts specific components (Stedmon et al. 2003), representing groups of fluorophores with similar fluorescence characteristics (Stedmon and Markager 2005). The model is defined as

$$x_{ijk} = \sum_{f=1}^{F} a_{if} b_{jf} c_{kf} + \varepsilon_{ijk}$$

$$i = 1, \dots, I; \quad j = 1, \dots, J; \quad k = 1, \dots, K$$

$$(1)$$

where  $x_{ijk}$  is the fluorescence intensity of the *i*th sample at emission wavelength j and excitation wavelength k.  $a_{if}$  is directly proportional to the concentration of the fth component in sample i.  $b_{if}$  is linearly related to the quantum efficiency (fraction of absorbed energy emitted as fluorescence) of the fth component at emission wavelength j.  $c_{kf}$  is linearly proportional to the specific absorption coefficient of the fth component at excitation wavelength k. The residual matrix  $\varepsilon_{ijk}$  represents the variability not accounted for by the model, and the model is found by minimising the sum of squared residuals (Stedmon et al. 2003).

The raw data were corrected for inner-filtering effects according to Parker and Barnes (1957). Inner filtering reduces the fluorescence intensity due to the absorption of the excitation beam and of emitted light after excitation (Lakowicz 1983, p. 44). The correction suggested by Lakowicz (1983, p. 44) and applied by e.g. McKnight et al. (2001) and Ohno (2002) gave nearly identical results as the Parker and Barnes (1957) correction, but the latter correction was selected because it takes account of the dimensions of the excitation and emission beams. The matrices were then corrected for instrument bias, i.e. the effects of lamp output and instrument sensitivity/ response. A blank, measured on a sealed cell of Milli-Q water, was subtracted to remove/reduce the Raman line (Hudson et al. 2007). Finally a mask was applied to the data matrix, setting all data in the regions without fluorescence (excitation wavelength exceeds emission wavelength) to zero, and replacing all data in the regions greatly influenced by the two Rayleigh-Tyndall lines (Hudson et al. 2007) with missing values (NaN in MATLAB).

A series of PARAFAC models with three to seven components were fitted to the data. As some of the components gave negative excitation or emission loadings, non-negativity constraints were applied. The PARAFAC model could be split half validated (i.e. separate modelling of two halves of the samples gave statistically identical results, cf. Stedmon and Bro 2008) for up to three components. Initial



exploration revealed that the sphagnum pore water samples from events 1 and 2 showed a high residual fluorescence at low excitation and emission wavelength. Some other samples, especially soil water samples, also exhibited some fluorescence in this region. However, overall this feature was present in too few samples for a model including a component in this region to be validated. Thus, the two sphagnum pore water samples were considered as outliers, and removed prior to the final modelling. The final model was the three component model giving the smallest residual error out of ten models fitted following random initialisation of the model. The maximum fluorescence intensity of the different components (Fmax) for individual samples was corrected for dilution by multiplying by the dilution factor.

#### Results

# Hydrology

The rainfall during events 1, 2 and 3 was 16, 17 and 114 mm, respectively (Fig. 2). Event 1 was the second in a series of events following an 11 day dry period. There was a 26 mm event just prior to event 1, and the stage was 198 mm at the onset of the

event. During the event, stage rose to 334 mm. Event 2 was preceded by 7 dry days, with a series of large events before that (total precipitation 347 mm). The stream stage was 138 mm at the onset of the event and maximum stage during the event was 211 mm. Event 3 was preceded by a 10 day dry period. The stream stage was at 131 mm at the onset. During the event, stage rose to 1,045 mm. Event 3 was clearly the largest of the three events, and one of the largest of the whole autumn.

## pH and DOC

The pH decreased during all events (Fig. 3). The largest decrease (2.3 pH units from base flow levels) occurred in event 3, compared to 0.9 and 1.1 pH units in event 1 and 2, respectively. Base flow pH was significantly higher than that of soil water in all events. pH of sphagnum pore water was at a similar level as that of soil water.

DOC concentrations increased during all events (Fig. 3), by 16, 44 and 89% relative to base flow in event 1, 2 and 3, respectively. The DOC concentration was significantly and negatively correlated with pH ( $r^2$  0.59, 0.81 and 0.86 for event 1, 2 and 3, respectively). Soil water DOC concentrations were higher than those of base flow, but the difference was only significant for events 2 and 3. DOC concentration in sphagnum pore water was higher than in soil

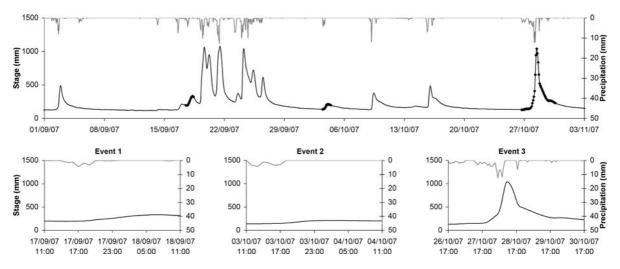
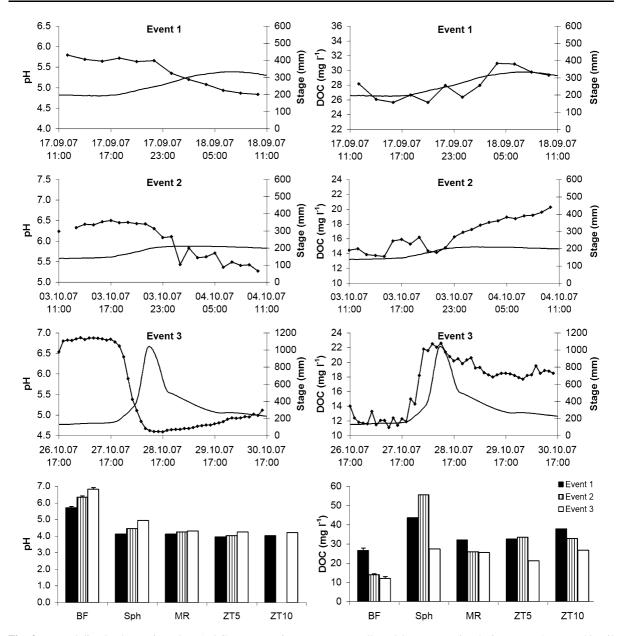


Fig. 2 Stream stage and precipitation for the whole autumn (top) and for single events (bottom). Markers in top panel indicate time of sampling. Note different time scale for event 3





**Fig. 3** pH and dissolved organic carbon (DOC) concentration in stream samples, sphagnum pore water and soil water from the three events. *BF* average base flow, *Sph* sphagnum pore water, *MR* soil water collected by micro-rhizons, *ZT5* soil

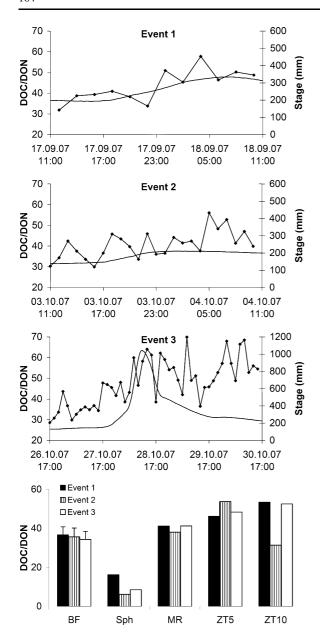
water collected by zero-tension lysimeter at 5 cm, *ZT10* soil water collected by zero-tension lysimeter at 10 cm. Note different time scale for event 3

water in events 1 and 2, and at the same level as in soil water in event 3.

## DOC/DON ratio

The DOC/DON ratio data (Fig. 4) were noisy, but there was an upward trend in all events. Maximum values were 58, 57 and 103% higher than base flow values in event 1, 2 and 3, respectively. There was no significant difference between soil water and base flow DOC/DON ratios in any of the events. The DOC/DON ratios of the sphagnum samples were far lower than those of both base flow and soil water samples.





**Fig. 4** Dissolved organic carbon/dissolved organic nitrogen (DOC/DON) ratio in stream samples, sphagnum pore water and soil water from the three events. *BF* average base flow, *Sph* sphagnum pore water, *MR* soil water collected by microrhizons, *ZT5* soil water collected by zero-tension lysimeter at 5 cm, *ZT10* soil water collected by zero-tension lysimeter at 10 cm. Note different time scale for event 3

#### Absorbance

Absorbance at 254 nm increased during events 2 and 3 (Fig. 5), with 30 and 53% maximum increases from base flow, respectively. In these events, absorbance

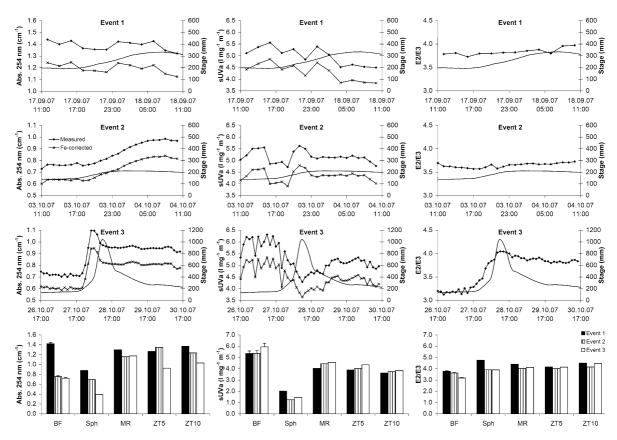
was significantly and positively correlated with DOC concentration ( $r^2$  0.88 and 0.92, respectively). In event 3 there was a striking diurnal pattern in absorbance after the initial peak, with consistent maxima at 7 am and minima at 1 pm. Soil water absorbance was significantly higher than that of base flow in events 2 and 3. Absorbance of sphagnum pore water was generally lower than that of both base flow and soil water.

The sUVa index (Fig. 5) decreased from base flow levels during event 1 (16% change), and more clearly during event 3 (28% change). There was no apparent pattern in event 2. The four deviating samples in event 2 with relatively low sUVa values correspond to the four samples with relatively low DOC, and appear to be caused by a DOC measurement error, as this deviation was not observed for absorbance. The reason for this error could not be established. Base flow sUVa was significantly higher than that of soil water in all events. sUVa in sphagnum pore water was far lower than sUVa in both base flow and soil water.

There was a rising trend in E2/E3 (Fig. 5) for all events, with a change from base flow to maximum levels of 5, 3 and 27% for event 1, 2 and 3, respectively. Soil water E2/E3 was significantly higher than that of base flow in all events. E2/E3 in sphagnum pore water was at a similar level as in soil water.

The average estimated Fe concentration was 2.2, 1.6 and 1.5 mg  $l^{-1}$  for event 1, 2 and 3, respectively, i.e. above the level  $(0.5 \text{ mg l}^{-1})$  where iron interference should be considered (Weishaar et al. 2003). The estimated Fe concentration followed closely the DOC concentration, and was only slightly modified by flow. Fig. 5 shows the effect of correcting for Fe absorbance on absorbance and sUVa. The effect is pronounced, and the resulting sUVa values are more consistent with previous research (cf. Weishaar et al. 2003, Table 1). However, the temporal patterns are preserved to a high degree. As the following discussion is mainly focused on the relative changes and temporal patterns, the issue of iron interference will not be further discussed, but it should be kept in mind that the absorbance and sUVa values are overestimated. E2/E3 is probably not strongly affected by iron interference. Weishaar et al. (2003) observed similar iron interference at 280 nm as 254 nm, so the interference at 365 nm is not believed to be very





**Fig. 5** Absorbance at 254 nm, sUVa (absorbance at 254 nm/DOC concentration) and E2/E3 (absorbance at 250 nm/absorbance at 365 nm) in stream samples, sphagnum pore water and soil water from the three events. *BF* average base

flow, *Sph* sphagnum pore water, *MR* soil water collected by micro-rhizons, *ZT5* soil water collected by zero-tension lysimeter at 5 cm, *ZT10* soil water collected by zero-tension lysimeter at 10 cm. Note different time scale for event 3

different. Moreover, the E2/E3 values obtained are consistent with previous research (cf. Peuravuori and Pihlaja 1997, Table 2).

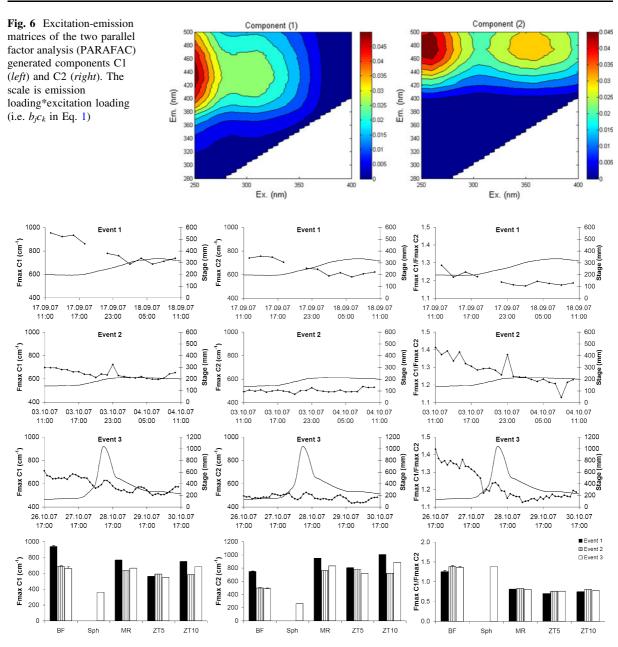
## Fluorescence

Component 3 from the PARAFAC analysis was considered to represent contamination, as it only appeared in the samples where a certain plastic tube was used for dilution. Blank samples using the same plastic tube exhibited the same feature. This component is consequently not discussed in the following. However, this component overlapped with the region that was poorly modelled for the sphagnum samples, and the presence of this contamination component may have added to the difficulty of validating a model with a true component in this region.

The two remaining components are shown in Fig. 6. Both components were double-peaked. Component 1 (C1) had maximum excitation at <250 nm with a secondary peak at 310 nm, and maximum emission at 440 nm. Component 2 (C2) had a maximum at excitation 260 nm with a secondary peak at 360 nm, and maximum emission at 474 nm.

The temporal variation in the fluorescence intensity (Fig. 7) did not seem to coincide with the changes in stream stage. Throughout the sampling periods the intensity levels were either stable or decreasing. C1 had the clearest decreasing trend. This resulted in a decrease in the C1/C2 ratio in all events, with a levelling off (events 1 and 2) or increase (event 3) towards the end of the events (Fig. 7). The decrease from base flow levels to minimum levels was 7, 18, and 17% in event 1, 2 and 3, respectively. C1 was higher in base flow than soil water, but this





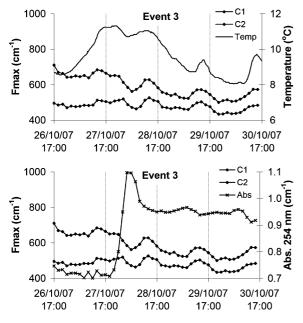
**Fig. 7** Maximum fluorescence intensity (Fmax) of component C1 and C2 and the ratio of the two in stream samples, sphagnum pore water and soil water from the three events. *BF* average base flow, *Sph* sphagnum pore water, *MR* soil water

collected by micro-rhizons, ZT5 soil water collected by zero-tension lysimeter at 5 cm, ZT10 soil water collected by zero-tension lysimeter at 10 cm. Note different time scale for event 3

was only significant for event 2. C2 was higher in soil water, and this was significant for events 2 and 3. Both C1 and C2 were lower in sphagnum pore water than in base flow and soil water. For C1/C2, base flow was significantly higher than soil water in all events. C1/C2 of sphagnum pore water was at the same level as for base flow.

In event 3, there was a clear diurnal pattern in C1 and C2, especially observed after the stage peak. The fluorescence peaked at 1 pm and was lowest during the night (9 pm to 7 am). The pattern coincided with the in situ temperature variation, while it was inversely related to the pattern observed for absorbance (Fig. 8).





**Fig. 8** Maximum fluorescence intensity (Fmax) of component C1 and C2 in event 3 plotted with in situ temperature (*top*) and absorbance at 254 nm (*bottom*)

Ohno et al. (2008) investigated Fe quenching of PARAFAC-derived components. Their results could be used as basis for a crude estimate of fluorescence quenching, as their components 1 and 2 corresponded well with C1 and C2, respectively. The effect on overall levels was pronounced, but the temporal patterns were preserved to a high degree. Ohno et al. (2008) showed that component 1 was more strongly quenched than component 2. However, the estimated effect of quenching on C1/C2 was minor. Hence, Fe quenching seems unimportant to the temporal patterns in fluorescence, and will not be further discussed.

# Mixing model

Mixing models for events 1 and 2 did not show any consistent patterns between parameters (data not shown). For event 3 (Fig. 9), there was a consistent, although not identical, pattern in H<sup>+</sup> concentration, DOC concentration and absorbance at 254 nm. The curves showed a decrease in the proportion of base flow-type water at the start of the event and a subsequent increase coinciding with the decrease in stream stage. The models based on H<sup>+</sup> and absorbance indicated lower and higher contribution of soil

water at peak flow compared to the model based on DOC concentration, respectively. Mixing models for event 3 based on C1 and C2 were not consistent with those of the other three parameters (Fig. 9).

#### Discussion

Effects of events on DOC concentration

The higher pH and lower DOC concentration in base flow compared to soil water indicates that the peat system in Afon Ddu is not like the system described by Clark et al. (2007, 2008), where base flow was chemically similar to soil water. Like DOC concentration, absorbance at 254 nm was also lower in base flow than in soil water. Absorbance is generally known to be closely correlated with DOC concentration (e.g. Dobbs et al. 1972; Brandstetter et al. 1996; Korshin et al. 1997). Relatively high baseflow DOC concentration indicates a significant input of drainage from the peat, whilst the high baseflow pH demonstrates some hydrological connectivity with underlying base-rich bedrock and/or mineral soil patches within the catchment.

The increased DOC concentration, increased absorbance and decreased pH at high flow can be explained by an increased contribution of soil water, due to a rising water table within the acrotelm and subsurface flow (Evans et al. 1999; Worrall et al. 2002). A higher similarity between soil water and stream water at high flow with respect to DOC concentrations was also observed by Billett et al. (2006). The change in stream chemistry could not solely be caused by an increased input of water from the sphagnum layer, as this could not have explained the trend in absorbance. There was little evidence of dilution at peak flow. The process could thus be reasonably well described for event 3 by the applied two end-member mixing model, rather than a three end-member model as suggested in Worrall et al. (2002). The discrepancies observed between the DOC concentration, H<sup>+</sup> concentration and absorbance at 254 nm were most likely due to the lack of conservative mixing. The most reliable model is probably the one for DOC, as H<sup>+</sup> concentration is unlikely to be conservative, and absorbance behaves abnormally at peak flow. The inconsistency between parameters in the models for events 1 and 2 can probably be



1200

1000

800

600

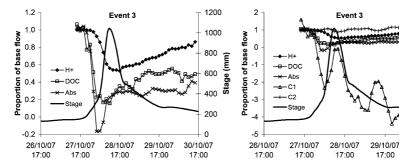
400

200

0

30/10/07

17:00



**Fig. 9** End-member mixing models based on different parameters in event 3. The *left panel* shows the models for H<sup>+</sup> concentration, DOC concentration and absorbance at 254 nm only, while the *right panel* also includes the mixing models based on C1 and C2. The mixing models show the proportion

of base flow type of water needed to explain stream concentration when the other end-member is soil water (average of the micro-rhizon sample and the two zero-tension lysimeter samples)

explained by the limited size of the events and small changes in stream chemistry, giving too much noise in such a crude model.

## Effects of events on DOM quality

The sUVa index provides a measure of the change in DOM quality expressed by deviation from the correlation between DOC concentration and absorbance. sUVa has been shown to be positively correlated with aromaticity and molecular weight (Croué et al. 1999; Weishaar et al. 2003; Hood et al. 2005). Both sUVa and aromaticity increase upon biodegradation (Kalbitz et al. 2003; Saadi et al. 2006). A high sUVa is also associated with lower bioavailability (Fellman et al. 2008). Thus, the decrease in sUVa at high flow indicates that the DOM released at high flow is less degraded and aromatic and of lower molecular weight. This is supported by the increase in E2/E3, as E2/E3 is negatively correlated to aromaticity and molecular weight (Peuravuori and Pihlaja 1997). E2/E3 is a more robust parameter than sUVa, giving a smoother trend.

DOC/DON data from this study are noisy, but clearly show increases in all three events. Increasing DOC/DON may be indicative of increasing aromaticity (McKnight et al. 1997; Hood et al. 2005), because algal and microbial material has low DOC/DON (McKnight et al. 1994) and low aromaticity (McKnight et al. 2001). However, even the lowest DOC/DON values observed are much higher than those associated with algal or microbial material (McKnight et al. 1994). Moreover, the highest DOC/

DON values observed correspond to the lower range of values observed for different litter leachates (Magill and Aber 2000). Hence, in this case it is more likely that increasing DOC/DON indicates decreasing degree of degradation (Melillo et al. 1982; Qualls and Haines 1992; Currie et al. 1996; Yano et al. 2004). Thus both absorbance and DOC/DON indicate that base flow is characterised by older and probably more degraded DOM, while soil water comprises younger, less degraded DOM (Evans et al. 2007).

With increased absorbance at high flow, one might expect to see a similar response in fluorescence. However, unlike absorbance, fluorescence does not invariably correlate well with DOC, and fluorescence relative to absorbance may vary between different sources (Baker et al. 2008). In addition, fluorescing structures constitute only a minor part of humic molecules (Miano et al. 1988; Senesi et al. 1991). Thus, it is possible for fluorescence to decrease even if absorbance, as a bulk parameter, increases.

The mixing model for event 3 indicates that stream fluorescence intensity during the event could not be described by a mixing of soil water and base flow water, as it could for absorbance. C2 remained approximately at base flow levels throughout the event despite the higher fluorescence intensity in soil water, giving a proportion of base flow water in the mixing model at about 1 all through the event. For C1 the proportion of base flow water was negative throughout most of the event, meaning that not even for the end-member with the lowest value (soil water), the fluorescence intensity was sufficiently low to cause the decrease in fluorescence intensity



observed during the event. The effect could not be explained by increased contribution of low fluorescence sphagnum pore water, as this does not fit with the observed changes in absorbance, DOC/DON and C1/C2 during the events. Rather, the lack of consistency between fluorescence parameters and the other parameters in the mixing models, the diurnal variation in fluorescence, and the co-variation in fluorescence and in situ stream temperature observed in event 3, collectively suggest that stream fluorescence may to a high degree be governed by in-stream microbial processes.

In-stream processing of DOM has been found to be an important control on stream DOC in another UK peatland catchment (Dawson et al. 2001). The microbial processing probably occurs in biofilms, as it is generally believed that this is where most microorganisms in natural systems exist (Sutherland 2001). Biofilms on the stream bed, rather than suspended aggregates in the water column tend to dominate ecosystems with high downstream transport and sediment-surface-area to water-volume ratio, as in headwaters (Battin et al. 2008). Lock and Hynes (1976) showed that the stream bottom, not the water itself is responsible for the major part of DOC removal, and biofilms on the stream bed have been shown to remove a substantial amount of DOC (Fiebig et al. 1990; Fiebig and Lock 1991). In the Afon Ddu there were no indications of in-stream processing of bulk DOC. This either confirms that only a minor part of the DOM released is subject to such processing, or that the spectrophotometric methods, introducing less uncertainty than the DOC analysis, are better for detecting short-term variation in DOM. The in-stream processing will be further covered in the next section.

Microbial processing of DOM is limited by microbial capacities, i.e. reaction rates (Battin et al. 2008). Hence, one can expect increased flow to give dilution of stream fluorescence. The trends in C1 and C2 can thus be explained by dilution as the main control, and only some compensation by fluorescing material from the soil. This indicates that highly fluorescing DOM in soil water is not as mobile as less fluorescing DOM. The compensation by soil water was highest for C2, where the levels were higher in soil water than in base flow. The C1/C2 ratio would not be affected by dilution, so the downwards trend was due to the small contribution of fluorescing material from the soil.

A decreased C1/C2 represents a shift in peak position to higher wavelengths, and this has been associated with higher density of aromatic rings, higher degree of aromatic substitution and conjugation, higher molecular weight, and higher hydrophobicity (Senesi 1990; Coble et al. 1998; Sharma and Schulman 1999, pp. 20-21; Wu et al. 2003). A peak shift to higher wavelengths may also signify an increase in the importance of humic compared to fulvic acids (Senesi 1990). The C1/C2 ratio thus suggests that the aromaticity and molecular weight of DOM was higher in soil water compared to base flow, which is opposite to the conclusion based on sUVa, E2/E3 and DOC/DON. However, this fits in well with the hypothesis that the fluorescing material is to a high degree produced in-stream (cf. discussion above and Dawson et al. 2001), which would make base flow fluorescing material relatively younger than that deriving from soil water, as opposed to what is the case for the bulk material. The overall conclusion is still that DOM released at high flow is generally less degraded, as the fluorescing part constitutes only a minor part of the DOM.

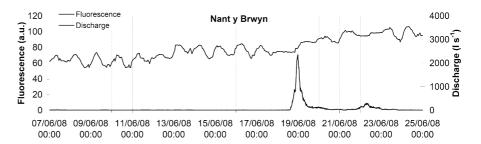
# Effects of events on in-stream processing of DOM

As discussed above, the diurnal pattern in fluorescence observed in event 3 appears to reflect diurnal variation in in-stream production of fluorescing DOM. Both temperature and light are environmental factors that vary on a daily timescale. Temperature has a positive effect on fluorescence, as the activity of the DOM-producing microorganisms is stimulated by increasing temperature (Christ and David 1996; Gödde et al. 1996; Freeman et al. 2001). UV light is known to cause a decrease in fluorescence, through so-called photo-bleaching (e.g. Skoog et al. 1996; Moran et al. 2000; Patel-Sorrentino et al. 2004), and can be expected to co-vary with stream temperature, peaking at mid-day.

To our knowledge a diurnal pattern in fluorescence has only been observed by Spencer et al. (2007). They measured in situ fluorescence during base flow by means of a fluorometer measuring at 370 nm excitation and over a broad emission band centred at 460 nm. However, whereas fluorescence in event 3 peaked at mid-day and was positively related to temperature, the diurnal pattern reported by Spencer et al. peaked in the early morning (around 9 am), had



Fig. 10 Fluorescence measured in situ (excitation 330 nm, emission 450 nm) and discharge in Nant y Brwyn



a minimum in late afternoon (around 6 pm), and was negatively related to temperature. According to Spencer et al., the diurnal pattern was related to day-time photo-bleaching of the DOM. This is confirmed by base flow data from similar in situ measurements (excitation 330 nm, emission 450 nm) from Afon Ddu (data not shown) and another stream (Nant y Brwyn), 2 km away on the same area of blanket peat (Fig. 10), which showed a similar diurnal pattern.

We hypothesise that during an event the diurnal fluorescence pattern is shifted, due to the extra input of soil water DOM represented by absorbing material. Nieto-Cid et al. (2006) showed that fluorescent humic substances could be produced by microbial degradation of DOM on a short (<1 day) timescale, and that the magnitude of production was related to microbial activity. If even a small proportion of the DOM transported from the soil can be used by biofilm microbial communities, this could explain the rapid, diurnal production of fluorescent DOM. Degradation would be expected to peak at mid-day due to the higher temperatures, which could explain the shift in the diurnal cycle, and also the negative relationship between fluorescence and absorbance in event 3. This positive effect of temperature apparently surpasses any negative effect of UV light at mid-day, perhaps because the cloud cover in the period during and after the storm event was high, and UV intensity consequently low. Alternatively, or additionally, given the high input of DOM, photo-bleaching could have had a positive impact on production of fluorescing DOM, as UV radiation is found to increase the substrate quality, and thus the degradability, of terrestrially derived DOM (Moran et al. 2000; Tranvik and Bertilsson 2001; Anesio et al. 2005).

The hypothesis appears to be supported by event data from the Nant y Brwyn (Fig. 10). At base flow the in situ fluorescence minimum was at about

4–7 pm and peaked in the early morning (6–8 am). In the first day of the event shown (June 18), the pattern was shifted, and after the peak in discharge (June 19) the in situ fluorescence remained high all day until a minimum at 8 pm. In the following few days in situ fluorescence continued to stay high throughout a longer part of the day, with a delayed minimum compared to base flow, until June 24, when the maximum was again at 8 am and the minimum at 4 pm. There was more noise in these data than the event data for the Afon Ddu, as the PARAFAC model involves smoothing. To compare, the intensities at the wavelength measured at Nant y Brwyn were extracted from the event 3 EEMs (data not shown), and although they exhibited more noise, the diurnal pattern could still be distinguished. If the hypothesis is correct, then at least part of the absorbing, humic DOM released during events must be rapidly processed close to where it was released. However, more data and a more quantitative approach are needed to confirm this. The hypothesis implies that the production of fluorescing DOM not only peaks at a different time of the day during and after an event, but that there is a general increase in fluorescence intensity. Dilution may explain why this was not observed.

## **Conclusions**

In peatlands where base flow is mainly high pH/low DOC ground water, events result in increased DOC concentration due to flushing of the shallow acrotelm. The increase in DOC is accompanied by decreased pH. The DOM released during events appears to be less degraded, less aromatic and more bioavailable. An increased input of this material is believed to have caused stimulation of in-stream microbial communities, producing fluorescing metabolites.



Absorbance proved to be a good indicator of changes in DOM quality and an identification of the shift of flow paths and sources throughout an event. Fluorescence was less useful in that respect, due to the apparent influence by in-stream production. However, absorbance and fluorescence data combined gave indications of short term changes in instream processing of DOM during events which have not previously been observed.

Acknowledgments This work was partly funded by the EU Eurolimpacs project (the Commission of European Communities GOCE-CT-2003-505540). We thank David Williams, Simon Grant, Annette Burden and Timothy Jones for assistance in the field; Annie Brittain and Steve Hughes for help with the chemical analyses; David Norris for drawing the catchment map; Colin Stedmon for assistance in the PARAFAC analysis; and professor Jan Mulder for valuable comments on the manuscript.

#### References

- Aitkenhead JA, McDowell WH (2000) Soil C:N ratio as a predictor of annual riverine DOC flux at local and global scales. Glob Biogeochem Cycles 14(1):127–138
- Aitkenhead JA, Hope D, Billett MF (1999) The relationship between dissolved organic carbon in stream water and soil organic carbon pools at different spatial scales. Hydrol Process 13(8):1289–1302
- Anesio AM, Graneli W, Aiken GR, Kieber DJ, Mopper K (2005) Effect of humic substance photodegradation on bacterial growth and respiration in lake water. Appl Environ Microbiol 71(10):6267–6275
- Baker A, Bolton L, Newson M, Spencer RGM (2008) Spectrophotometric properties of surface water dissolved organic matter in an afforested upland peat catchment. Hydrol Process 22(13):2325–2336
- Battin TJ, Kaplan LA, Findlay S, Hopkinson CS, Marti E, Packman AI, Newbold JD, Sabater F (2008) Biophysical controls on organic carbon fluxes in fluvial networks. Nat Geosci 1(2):95–100
- Billett MF, Deacon CM, Palmer SM, Dawson JJC, Hope D (2006) Connecting organic carbon in stream water and soils in a peatland catchment. J Geophys Res 111(G2): G02010. doi:10.1029/2005JG000065
- Billett MF, Garnett MH, Harvey F (2007) UK peatland streams release old carbon dioxide to the atmosphere and young dissolved organic carbon to rivers. Geophys Res Lett 34(23):L23401. doi:10.1029/2007GL031797
- Boyer EW, Hornberger GM, Bencala KE, McKnight DM (1997) Response characteristics of DOC flushing in an alpine catchment. Hydrol Process 11(12):1635–1647
- Brandstetter A, Sletten RS, Mentler A, Wenzel WW (1996) Estimating dissolved organic carbon in natural waters by UV absorbance (254 nm). J Plant Nutr Soil Sci 159(6): 605–607

- Buffam I, Galloway JN, Blum LK, McGlathery KJ (2001) A stormflow/baseflow comparison of dissolved organic matter concentrations and bioavailability in an Appalachian stream. Biogeochemistry 53(3):269–306
- Christ MJ, David MB (1996) Temperature and moisture effects on the production of dissolved organic carbon in a Spodosol. Soil Biol Biochem 28(9):1191–1199
- Christensen JH, Hewitson B, Busuioc A, Chen A, Gao X, Held I, Jones R, Kolli RK, Kwon W-T, Laprise R, Magaña Rueda V, Mearns L, Menéndez CG, Räisänen J, Rinke A, Sarr A, Whetton P (2007) Regional climate projections. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL (eds) Climate change 2007: the physical science basis. Contribution of working group I to the fourth assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge
- Clark JM, Lane SN, Chapman PJ, Adamson JK (2007) Export of dissolved organic carbon from an upland peatland during storm events: implications for flux estimates. J Hydrol 347(3–4):438–447
- Clark JM, Lane SN, Chapman PJ, Adamson JK (2008) Link between DOC in near surface peat and stream water in an upland catchment. Sci Total Environ 404(2–3):308–315
- Coble PG, Del Castillo CE, Avril B (1998) Distribution and optical properties of CDOM in the Arabian Sea during the 1995 Southwest Monsoon. Deep-Sea Res Part II 45(10–11):2195–2223
- Countryside Council for Wales (2008) Yr Wyddfa—Snowdon ECN Site. https://weather.ccw.gov.uk/. Cited 25 Jan 2008
- Croué J-P, Violleau D, Bodaire C, Legube B (1999) Removal of hydrophobic and hydrophilic constituents by anion exchange resin. Water Sci Technol 40(9):207–214
- Currie WS, Aber JD, McDowell WH, Boone RD, Magill AH (1996) Vertical transport of dissolved organic C and N under long-term N amendments in pine and hardwood forests. Biogeochemistry 35(3):471–505
- Dawson JJC, Bakewell C, Billett MF (2001) Is in-stream processing an important control on spatial changes in carbon fluxes in headwater catchments? Sci Total Environ 265(1–3):153–167
- Dobbs RA, Dean RB, Wise RH (1972) Use of ultraviolet absorbance for monitoring total organic carbon content of water and wastewater. Water Res 6(10):1173–1180
- Eimers MC, Buttle J, Watmough SA (2008) Influence of seasonal changes in runoff and extreme events on dissolved organic carbon trends in wetland- and upland-draining streams. Can J Fish Aquat Sci 65(5):796–808
- Evans MG, Burt TP, Holden J, Adamson JK (1999) Runoff generation and water table fluctuations in blanket peat: evidence from UK data spanning the dry summer of 1995. J Hydrol 221(3-4):141–160
- Evans CD, Freeman C, Cork LG, Thomas DN, Reynolds B, Billett MF, Garnett MH, Norris D (2007) Evidence against recent climate-induced destabilisation of soil carbon from <sup>14</sup>C analysis of riverine dissolved organic matter. Geophys Res Lett 34(7):L07407. doi:10.1029/2007 GL029431
- Fellman JB, D'Amore DV, Hood E, Boone RD (2008) Fluorescence characteristics and biodegradability of dissolved organic matter in forest and wetland soils from coastal



- temperate watersheds in southeast Alaska. Biogeochemistry 88(2):169-184
- Fiebig DM, Lock MA (1991) Immobilization of dissolved organic matter from groundwater discharging through the stream bed. Freshw Biol 26(1):45–55
- Fiebig DM, Lock MA, Neal C (1990) Soil water in the riparian zone as a source of carbon for a headwater stream. J Hydrol 116(1–4):217–237
- Freeman C, Evans CD, Monteith DT, Reynolds B, Fenner N (2001) Export of organic carbon from peat soils. Nature 412(6849):785
- Gödde M, David MB, Christ MJ, Kaupenjohann M, Vance GF (1996) Carbon mobilization from the forest floor under red spruce in the northeastern USA. Soil Biol Biochem 28(9):1181–1189
- Hinton MJ, Schiff SL, English MC (1997) The significance of storms for the concentration and export of dissolved organic carbon from two Precambrian Shield catchments. Biogeochemistry 36(1):67–88
- Hinton MJ, Schiff SL, English MC (1998) Sources and flowpaths of dissolved organic carbon during storms in two forested watersheds of the Precambrian Shield. Biogeochemistry 41(2):175–197
- Hood E, Williams MW, McKnight DM (2005) Sources of dissolved organic matter (DOM) in a Rocky Mountain stream using chemical fractionation and stable isotopes. Biogeochemistry 74(2):231–255
- Hope D, Billett MF, Cresser MS (1994) A review of the export of carbon in river water—fluxes and processes. Environ Pollut 84(3):301–324
- Hudson N, Baker A, Reynolds D (2007) Fluorescence analysis of dissolved organic matter in natural, waste and polluted waters—A review. River Res Appl 23(6):631–649
- Inamdar SP, O'Leary N, Mitchell MJ, Riley JT (2006) The impact of storm events on solute exports from a glaciated forested watershed in western New York, USA. Hydrol Process 20(16):3423–3439
- Kalbitz K, Schwesig D, Schmerwitz J, Kaiser K, Haumaier L, Glaser B, Ellerbrock R, Leinweber P (2003) Changes in properties of soil-derived dissolved organic matter induced by biodegradation. Soil Biol Biochem 35(8):1129–1142
- Korshin GV, Li CW, Benjamin MM (1997) Monitoring the properties of natural organic matter through UV spectroscopy: a consistent theory. Water Res 31(7):1787–1795
- Lakowicz JR (1983) Principles of fluorescence spectroscopy. Plenum Press, New York
- Lock MA, Hynes HBN (1976) The fate of "dissolved" organic carbon derived from autumn-shed maple leaves (Acer saccharum) in a temperate hard-water stream. Limnol Oceanogr 21(3):436–443
- Magill AH, Aber JD (2000) Dissolved organic carbon and nitrogen relationships in forest litter as affected by nitrogen deposition. Soil Biol Biochem 32(5):603–613
- McDowell WH, Likens GE (1988) Origin, composition, and flux of dissolved organic carbon in the Hubbard Brook valley. Ecol Monogr 58(3):177–195
- McKnight DM, Andrews ED, Spaulding SA, Aiken GR (1994) Aquatic fulvic acids in algal-rich antarctic ponds. Limnol Oceanogr 39(8):1972–1979
- McKnight DM, Harnish R, Wershaw RL, Baron JS, Schiff S (1997) Chemical characteristics of particulate, colloidal,

- and dissolved organic material in Loch Vale Watershed, Rocky Mountain National Park. Biogeochemistry 36(1): 99–124
- McKnight DM, Boyer EW, Westerhoff PK, Doran PT, Kulbe T, Andersen DT (2001) Spectrofluorometric characterization of dissolved organic matter for indication of precursor organic material and aromaticity. Limnol Oceanogr 46(1):38–48
- Melillo JM, Aber JD, Muratore JF (1982) Nitrogen and lignin control of hardwood leaf litter decomposition dynamics. Ecology 63(3):621–626
- Miano TM, Sposito G, Martin JP (1988) Fluorescence spectroscopy of humic substances. Soil Sci Soc Am J 52(4):1016–1019
- Moran MA, Sheldon WM, Zepp RG (2000) Carbon loss and optical property changes during long-term photochemical and biological degradation of estuarine dissolved organic matter. Limnol Oceanogr 45(6): 1254–1264
- Neal C, Lofts S, Evans CD, Reynolds B, Tipping E, Neal M (2008) Increasing iron concentrations in UK upland waters. Aquat Geochem 14(3):263–288
- Nieto-Cid M, Álvarez-Salgado XA, Pérez FF (2006) Microbial and photochemical reactivity of fluorescent dissolved organic matter in a coastal upwelling system. Limnol Oceanogr 51(3):1391–1400
- Ohno T (2002) Fluorescence inner-filtering correction for determining the humification index of dissolved organic matter. Environ Sci Technol 36(4):742–746
- Ohno T, Amirbahman A, Bro R (2008) Parallel factor analysis of excitation-emission matrix fluorescence spectra of water soluble soil organic matter as basis for the determination of conditional metal binding parameters. Environ Sci Technol 42(1):186–192
- Parker CA, Barnes WJ (1957) Some experiments with spectrofluorimeters and filter fluorimeters. Analyst 82(978): 606–618
- Patel-Sorrentino N, Mounier S, Lucas Y, Benaim JY (2004) Effects of UV-visible irradiation on natural organic matter from the Amazon basin. Sci Total Environ 321(1–3): 231–239
- Peuravuori J, Pihlaja K (1997) Molecular size distribution and spectroscopic properties of aquatic humic substances. Anal Chim Acta 337(2):133–149
- Qualls RG, Haines BL (1992) Biodegradability of dissolved organic matter in forest throughfall, soil solution, and stream water. Soil Sci Soc Am J 56(2):578–586
- Saadi I, Borisover M, Armon R, Laor Y (2006) Monitoring of effluent DOM biodegradation using fluorescence, UV and DOC measurements. Chemosphere 63(3):530–539
- Senesi N (1990) Molecular and quantitative aspects of the chemistry of fulvic acid and its interactions with metal ions and organic chemicals: Part II. The fluorescence spectroscopy approach. Anal Chim Acta 232(1):77–106
- Senesi N, Miano TM, Provenzano MR, Brunetti G (1991) Characterization, differentiation, and classification of humic substances by fluorescence spectroscopy. Soil Sci 152(4):259–271
- Sharma A, Schulman SG (1999) Introduction to fluorescence spectroscopy. Wiley, New York



- Skoog A, Wedborg M, Fogelqvist E (1996) Photobleaching of fluorescence and the organic carbon concentration in a coastal environment. Mar Chem 55(3–4):333–345
- Soulsby C, Rodgers P, Smart R, Dawson J, Dunn S (2003) A tracer-based assessment of hydrological pathways at different spatial scales in a mesoscale Scottish catchment. Hydrol Process 17(4):759–777
- Spencer RGM, Pellerin BA, Bergamaschi BA, Downing BD, Kraus TEC, Smart DR, Dahgren RA, Hernes PJ (2007) Diurnal variability in riverine dissolved organic matter composition determined by in situ optical measurement in the San Joaquin River (California, USA). Hydrol Process 21(23):3181–3189
- Stedmon CA, Bro R (2008) Characterizing dissolved organic matter fluorescence with parallel factor analysis: a tutorial. Limnol Oceanogr Methods 6:572–579
- Stedmon CA, Markager S (2005) Resolving the variability in dissolved organic matter fluorescence in a temperate estuary and its catchment using PARAFAC analysis. Limnol Oceanogr 50(2):686–697
- Stedmon CA, Markager S, Bro R (2003) Tracing dissolved organic matter in aquatic environments using a new approach to fluorescence spectroscopy. Mar Chem 82(3–4):239–254
- Sutherland IW (2001) The biofilm matrix—an immobilized but dynamic microbial environment. Trends Microbiol 9(5):222–227
- Thurman EM (1985) Organic geochemistry of natural waters. Nijhoff, Dordrecht
- Tranvik LJ, Bertilsson S (2001) Contrasting effects of solar UV radiation on dissolved organic sources for bacterial growth. Ecol Lett 4(5):458–463

- Vogt RD, Akkanen J, Andersen DO, Bruggemann R, Chatterjee B, Gjessing E, Kukkonen JVK, Larsen HE, Luster J, Paul A, Pflugmacher S, Starr M, Steinberg CEW, Schmitt-Kopplin P, Zsolnay A (2004) Key site variables governing the functional characteristics of dissolved natural organic matter (DNOM) in Nordic forested catchments. Aquat Sci 66(2):195–210
- Weishaar JL, Aiken GR, Bergamaschi BA, Fram MS, Fujii R, Mopper K (2003) Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. Environ Sci Technol 37(20):4702–4708
- Worrall F, Burt TP, Jaeban RY, Warburton J, Shedden R (2002) Release of dissolved organic carbon from upland peat. Hydrol Process 16(17):3487–3504
- Wu FC, Evans RD, Dillon PJ (2003) Separation and characterization of NOM by high-performance liquid chromatography and on-line three-dimensional excitation emission matrix fluorescence detection. Environ Sci Technol 37(16): 3687–3693
- Yano Y, Lajtha K, Sollins P, Caldwell BA (2004) Chemical and seasonal controls on the dynamics of dissolved organic matter in a coniferous old-growth stand in the Pacific Northwest, USA. Biogeochemistry 71(2):197–223
- Zepp RG, Sheldon WM, Moran MA (2004) Dissolved organic fluorophores in southeastern US coastal waters: correction method for eliminating Rayleigh and Raman scattering peaks in excitation-emission matrices. Mar Chem 89(1–4):15–36

